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# Antioxidant and Antidiabetic Activities of Ethanolic Leaf Extract of *Melochia corchorifolia*

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#### **Abstract**

Polyphenolic extract of Melochia corchorifolia leaf has demonstrated high antioxidative activity and very promising anti-diabetic activities in vitro. The antioxidant and antidiabetic inhibition properties of the polyphenolic-rich extracts were also studied in this study and it revealed that the plant extract showed better activities than the conventional antidiabetic drug acarbose. The biological activity of Melochia corchorifolia leaf extracts was investigated. The maximum inhibition for the DPPH inhibition assay of Melochia corchorifolia was recorded at 10 μg/ml, where the scavenging activity of the control drug (Ascorbic acid) was found to be better than that of the plant extract at all dosages except for 20 µg/ml and 50 µg/ml, where Melochia corchorifolia showed better activity than the control. The ABTS inhibition assay result showed that the activity of the extract and the control is dose-dependent. The ethanolic leaf extract of Melochia corchorifolia demonstrated significant antioxidant (DPPH IC50: 66.63 μg/mL; ABTS: 87.37 mM TE/g) and antidiabetic (α-amylase inhibition: 68.62 %) activities, surpassing acarbose in  $\alpha$ -glucosidase inhibition (p < 0.05). The activity increases with an increase in the dosage. The best activity was recorded at the lowest dosage i.e., 10 µg/ml, where the plant extract inhibition activity was (6.474) at the same dosage against (27.394) of the positive control. This plant may hold the potential against diabetes.

Keywords: Melochia corchorifolia; Ascorbic acid; anti-diabetic and antioxidant

#### Introduction

The occurrence of bioactive plant components, mainly called phytochemicals, has been considered of fundamental nutritional importance in the prevention of many diseases such as cancer, cardiovascular disease, and diabetes (Aruoma, 2003). It has been discovered that regular utilization of fruits, vegetables, herbs, and spices has been associated with health benefits for humans and animals. But not until recently, these valuable compounds (phytochemistry and biological activity) were discovered to possess a wide range of biological activity (Sheetal and Jamuna, 2009). These biological activities of plants help in health benefits beyond basic nutrition (Oomah and Mazza, 2008).

Hence, biological activities studies of medicinal plants that are common in northern Nigeria will go a long way in providing a cheaper way of treating diseases such as jaundice, intestinal worm infection, wounds, malaria, venereal diseases, epilepsy, diarrhea, hemorrhoid, cancer, asthma and fever afflicting substantial number of communities in the region. Natural products are chemical organic substances that are produced by living organisms found in nature through the pathways of secondary and primary metabolisms (Woldeyes *et al.*, 2012).

Natural products have pharmacological activities that can be useful in treating various diseases, and may act as active substances for both modern medicines and traditional medicine (Raafat, 2013). Medicinal plants are plants that have at least one of their parts roots, stems, barks, or leaves, used for therapeutic purposes (*Chinyere et al.*, 2011).

#### MATERIALS AND METHODS

#### **Chemicals / Reagent**

All the chemicals and reagents used in the studies were of analytical grade and obtained from the Federal University Dutsin-Ma, Katsina State, in the Department of Applied Chemistry. Dichloromethane, n-Hexane, ethyl acetate, ethanol, concentrated sulfuric acid, chloroform, 5% acetone, hydrochloric acid, acetic anhydride, distilled water, disinfectant, amongst others were used in the work.

## **Equipment**

Rotatory evaporator (Bibby Scientific Limited, Stone Staffordshire, ST15 OSA.UK), Vacuum pump, water bath, electric shaker. Analytical balance, Top loading balance, Hot water bath and ultraviolet light (254nm & 366nm).

#### Methods

#### **Collection of Plant Material**

During the rainy season in September 2022, the leaves of *Melochia corchorifolia* were collected from Wakaji village in the Dutsin-Ma local government of Katsina state, Nigeria. It was discovered at the Federal University of Dutsin-Ma Katsina, Department of Plant Science. The leaf was

cleaned with water and air-dried for two weeks in the laboratory. Pestle and mortar were used to grind the leaf. The powdered samples were kept in clean, airtight containers at room temperature until they were needed.

#### **Extraction of Plant Materials**

The powdered sample (1.2 kg) was packed in a speculator and was extracted (Maceration) successively using ethanol (maceration). At the end of the extraction, the extracts obtained were concentrated at 40 °C using a rotatory evaporator, dried to a constant weight in a pre-weighed petri-dish and then kept in a refrigerator before subsequent analysis.

#### **Antioxidant Activity**

# 1-1-dipheny l-2 picrylhydrazyl free radical scavenging activity (DPPH):

The free radical scavenging ability of the extracts against DPPH (1, 1- diphenyl–2 picrylhydrazyl) free radical was evaluated using a modified method as described by Tuba and Gulcin (2008). Briefly, a 0.3 mm solution of DPPH was prepared in methanol and 500  $\mu$ L of the DPPH solution was added to 1 mL of the extracts at various concentrations (15–240  $\mu$ g/mL). These solutions were mixed and incubated in the dark for 30 minutes at room temperature. The absorbance was read at 517 nm against blank samples lacking a scavenger.

# Azino-bis(3-ethylbenzothiazoline-6-sulphonic acid) (ABTS) scavenging activity of ABTS scavenging activity:

The ABTS scavenging activity of the plant extract was determined using different

methods of Analysis were carried out in triplicate.

## Alpha amylase inhibitory activity

The  $\alpha$ -amylase inhibitory activity was measured according to (Shai et al., 2010) with slight modifications. A volume of 250 uL of each extract or acarbose at different concentrations (100-500) $\mu g/mL$ ) incubated with 500 µL of porcine pancreatic amylase (2µ/mL) in phosphate buffer (100 mM, pH 6.8) at 37 °C for 20 minutes. Then, 250 µL of 1% starch dissolved in 100 mM phosphate buffer (pH 6.8) was further added to the reaction mixture and then incubated at 37 °C for 1 hour. Then, dinitro salicylate color reagent (1 mL) was added then left to boil for 10 minutes. The mixture resulted in an absorbance that read at 540nm, and the inhibitory activity was expressed without the inhibitors as a percentage of the control.

# Alpha-glucosidase inhibitory activity

The  $\alpha$ -glucosidase inhibitory activity was determined by the method described by (Ademiluyi and Oboh, 2013) with slight modifications. Briefly, 250 µL of each acarbose different extract or at concentrations (30-240) $\mu g/mL$ ) was incubated with 500  $\mu L$  of 1.0 U/mL  $\alpha$ glucosidase solution in 100 mM phosphate buffer (100mM, pH 6.8) at 37 °C for 15 minutes. Then, 250 µL of pNPG solution (5 mM) in phosphate buffer (100 mM, pH 6.8) was added and the reaction mixture was further incubated at 37 °C for 20 minutes. The absorbance of the released pnitrophenol was measured at 405 nm and the inhibitory activity was expressed as a percentage of the control without the

inhibition. All assays were carried out in triplicate. The inhibitory activities of the extracts on the  $\alpha$ -glucosidase and  $\alpha$ -amylase were calculated by using the following formula:

Inhibitory activity  $\% = (1 - as/Ac) \times 100$ 

Where As: is the absorbance in the presence of the sample and

Ac: is the absorbance of the control.

The concentration resulting in inhibition of 50% enzyme activity and scavenging activities (IC50) weas obtained by calculating from the plot of percentage inhibition against log (concentration of the samples).

#### RESULTS AND DISCUSSION

# EXTRACTION OF Melochia corchorifolia

Cold maceration extraction was used to extract 1000g of the *M. corchorifolia* using bottles. Hexane was initially used, followed by dichloromethane, and ethanol, respectively. Each of the crude extracts was concentrated by using a rotary evaporator, and the resulting crude extracts were airdried until constant weights were obtained. The yield of crude methanol extract (0.56%) is the highest among the four samples, whereas the yield of crude dichloromethane extract (0.34 %) is the lowest compared to the other samples.

Yield % = weight of dry extract/weight of dry plant × 100%

Yield % = weight of dry extract/weight of dry plant  $\times$  100%

**Table 1: Extraction Yield** 

SOLVENTS	Weight of extract (g)	PERCENTAGE	YIELD
		(%)	
n-hexane	4.00	0.4	
Dichloromethane	3.60	0.34	
Ethanol	6.00	0.56	

#### Biological Activity of M. corchorifolia

Table 2: DPPH inhibition assay of Ethanol fraction

Dosage	10 μg/ml	20 μg/ml	50 μg/ml	100 μg/ml	150 μg/ml
PFMC	31.67	40.03	46.99	62.12	66.63
ASC	30.07	41.82	51.97	60.81	65.41

Table 2 shows that the activity of the plant extract in DPPH solution is dose-dependent. The positive control used is ascorbic acid, which is the conventional drug against oxidative stress. From the table, maximum inhibition was recorded at 10 μg/ml where the scavenging activity of the control drug (Ascorbic acid) was found to be better than that of the plant extracts at all dosages. *M. corchorifolia* showed better activity than the control. In general, the scavenging activity of both the control and the plant extract increases

with a corresponding increase in dosage. The ABTS inhibition assay result showed that the activity of the extract and the control is dose-dependent. The activity increases with a decrease in the concentration. The best activity was recorded at the highest dosage i.e., 150 μg/ml. *M. corchorifolia* has the best activity at (6.474), whose inhibition activity was at the same dosage against (27.39) of the positive control. The plant extracts showed better activity than the control at all concentrations.

**Table 3: ABTS Inhibition Assay of Ethanol Fraction** 

Dosage	10 μg/ml	20 μg/ml	50 μg/ml	100	150 μg/ml
				μg/ml	
PFMC	6.47	31.14	56.82	87.37	87.37
ASC	27.39	55.80	78.2 2	86.35	90.10

Table 4: H<sub>2</sub>O<sub>2</sub> Inhibition Assay of Ethanol Fraction

Dosage	10 μg/ml	20 μg/ml	<i>50</i> μg/ml	100 μg/ml	150 μg/ml
PFMC	13.89	24.44	34.28	47.32	52.92

Table 5: α-amylase Inhibition Assay of Ethanol Fraction

Dosage	20 μg/ml	50 μg/ml	100 μg/ml	250 μg/1	ml 500 μf/ml
PFMC	29.90	40.52	29.41	21.07	17.15
ASC	42.48	58.00	63.88	67.15	68.62

Table 6: α-glucosidase Inhibition Assay of Ethanol Fraction

Dosage	20μg/ml	50μg/ml	100μg/ml	250μg/ml	500µg/ml	
PFMC	22.77	24.20	24.74	26.52	28.74	
ASC	90.33	93.27	93.700	93.72	94.03	

Table 7: Advanced Glycation End-product assay

Dosage	M. corchorifolia
1mg/ml	24.84726

Results from the H<sub>2</sub>O<sub>2</sub> inhibition assay showed that the activity of the plant extracts is dose-dependent. This result also follows a similar trend to other antioxidant assays studied in the research. Maximum activity was recorded at the lowest dosage, 150 µg/ml, where the scavenging activity of ascorbic acid was the best. The antidiabetic potential of the leaf extract of M. corchorifolia was studied. The result showed that the activity of the plant extracts is dose-dependent. The activity increases with increasing dosage of the extracts. The best activity was recorded at a dosage of 500  $\mu$ g/ml. Table 6 shows the  $\alpha$ -glucosidase activity of M. corchorifolia. The table shows the inhibition activity that of corchorifolia leaves extract is dosedependent. There is an increase in the activity of both polyphenolic-rich extracts with a corresponding increase in dosage, with optimum activity achieved at a dosage of 500  $\mu$ g/ml. In general, the activity of M. *Melochia* was the best at the optimum dosage. Table 7 shows the advanced glycation end-products inhibition of *M. corchorifolia*, the plant extract showed activity on the AGE inhibition assay with an activity of 75.1% in *Melochia corchorifolia*.

### **Antioxidant activity**

Oxidative stress and viral disease have long been regarded as major causes of morbidity and mortality in humans. In order to achieve stability, free radicals destroy other molecules by removing electrons from them. The adverse effects of routinely used conventional antioxidants have compelled the quest for natural antioxidants from plant extracts (Tsado *et al.*, 2016). The DPPH radicals were frequently utilized to study the scavenging activity of various natural substances. In a DPPH radical scavenging

extract leaf assay, ethanol of M. was have corchorifolia found to a substantial antioxidant effect, with maximal inhibition of 34.1429 at 10 ug/ml. The decrease in DPPH absorbance generated by M. Corchorifolia polyphenolic-rich fraction is due to the reaction between antioxidant molecules and radicals, which results in radical scavenging via hydrogen Natural products donation. contain numerous antioxidant components. Flavonoids are phenolic chemicals that play essential roles in scavenging free radicals and consequently play critical functions in the prevention of oxidative stress-related disorders (Nahak and Sahu, 2010).

This strong scavenging property was related to the presence of hydroxyl groups in the chemical structure of phenolic compounds, which can offer the required components as a radical scavenger. These findings back up previous observations that plant metabolites such as phenol, tannins, and flavonoids have antioxidant and antibacterial activities. The DPPH antioxidant result obtained in this study goes hand in hand with the result obtained by Purushotham et al. (2019), where the control (Ascorbic acid) has slightly better activity than that of the plant extract. In another study conducted by Rao et al. (2012). M. corchorifolia showed good antioxidant activity in DPPH, Peroxide and superoxide assay with IC<sub>50</sub> values of 179  $\mu g/ml$ , 240 μg/ml and  $127\mu g/ml$ , respectively.

The antioxidant activity of plant extracts is measured using the ABTS radical. The radical absorbs at 743nm to produce a bluish green color, and an electron is lost by the nitrogen atom of ABTS, resulting in the production of the radical (Abdulrahman *et* 

al., 2021). Both plants employed in this study showed superior activity against the ABTS solution than the control (Ascorbic acid), this may be due to both of them being leaves and having high volume of polyphenol in them and there is a link between polyphenolic content and antioxidant activity, this is in tandem with the finding of Adegbola et al., 2020 where the antioxidant activity of A. hybridus was related to polyphenolic chemicals detected in significant concentrations in the plant's methanol leaf extract.

#### **Antidiabetic activity**

Diabetes is a chronic condition of carbohydrate, protein, and lipid metabolism caused by an absolute or relative lack of insulin secretion with/without variable degrees of insulin resistance (Barar, 2000). It can also be characterized as a disease in which the body either produces little or no insulin, or grows increasingly resistant to its activity (Ranian and Ramanujam, 2002). It is now an epidemic, with a global frequency of 5% in the general population. The global adult diabetes population will increase from 135 million in 1995 to 300 million by 2025 2002). Diabetes mellitus is (Torben, probably the fastest-growing metabolic disease in the world and as knowledge of the heterogeneous nature of the increases, so does the need for more challenging and appropriate therapies. Traditional plant remedies have been used for centuries in the treatment of diabetes, but only a few have been scientifically evaluated (Kumar et al., 2012). In the alpha-amylase assay, M. Corchorifolia exhibited excellent inhibition activity, where the activity of the increased extract with increasing

concentration or dosage. This is supported by the findings of research carried out (Vinoth et al., 2021), where Melochia corchorifolia inhibition is highly demonstrated in the enzyme at all concentrations. Plant phenolic and flavonoid compounds such as quercetin, ferulic acid, anthocyanins, catechin, and resveratrol were shown in epidemiological studies to regulate glycemia via increased glucose uptake, insulin secretion, and inhibition of lipid peroxidation, alphaglucosidase, and alpha-amylase (Lin et al., 2016). Studies have demonstrated that flavonols and flavones' enzyme inhibitory capacity depends on hydrogen bonds between the hydroxyl groups of the polyphenol ligands and the catalytic residues of the binding site and formation of a conjugated α-system that stabilizes the interaction with the active site (Xiao et al., 2015). The recent findings show structure-activity relationship of polyphenols inhibiting α-amylase have concluded that the hydroxylation of including galloylation flavonoids, catechins, improved the inhibitory effects against α-amylase (Wong et al., 2015). In the  $\alpha$ -glucosidase and  $\alpha$ -amylase antidabetic assays carried out in this study, the two plant samples compete favourably compared to that observed for the control drug acarbose employed in this study. To further buttress this claim, Yang et al., (2020) reported that the water extracts of A. gangeticus and A. inamoenus showed high α-glucosidase inhibitory activity (80-50%) at a concentration of 1 mg. Another study by (Mondal et al., 2015), the methanol extract Melochia corchorifolia significant α-glucosidase inhibitory activity

(IC50 8.49  $\mu$ M/ml) and that acarbose provides inhibition at IC<sub>50</sub> values of 15.25  $\mu$ M/ml.

#### Conclusion

In conclusion, the polyphenolic extract of *M. corchorifolia* leaves has demonstrated high antioxidative activity and very promising anti-diabetic activities *in vitro* with better activity. The antioxidant and antidiabetic inhibition properties of the polyphenolic-rich extracts were also studied, and it revealed that the plant extract showed better activities than the conventional antidiabetic drug acarbose.

#### Reference

Adebooye, O.C. 1996. Proximate composition and nutrient analyzes of six selected leaf vegetables of Southwest Nigeria. Ife Journal of Agriculture, 18 (1,2): 56-62.

Adebooye, O.C. 2000. An assessment of cultural practices for cultivating a wild but edible leaf vegetable: Crassocephalum biafrae (Asteraceae): Emphasis on propagation techniques. Proceedings of the In: Third International Workshop on the Sustainable Use of Medicinal and Food Plants. Karachi, Pakistan. pp. 132-138.

Adebooye, O.C. 2001. Wild plants for medicinal and culinary use: Nigeria. In: Sharing innovative experience on sustainable use of indigenous food and medicinal plants. Third World Academy of Sciences and UNDP, Trieste, Italy. pp. 6978.

- Adebooye, O.C., 2000. An assessment of cultural practices for cultivating a wild but edible leaf vegetable: Crassocephalum biafrae (Asteraceae): Emphasis on propagation techniques. Proceedings of In: the third international workshop on the sustainable use of medicinal and food plants. September 15–17, 2000. University of Karachi, Karachi, Pakistan. pp. 132–138.
- Adedayo, B.C., Oboh, G., Oyeleye, S.I., Ejakpovi, I.I., Boligon, A.A., & Athayde, M.L. Blanching alters the phenolic constituents and in vitro antioxidant and anticholinesterases properties of fireweed (Crassocephalum crepidioides). Journal of Taibah University Medical Sciences, 10(4): 419–426, (2015).
- Adelakun SA, Ogunlade B, Omotoso OD, Oyewo OO. Role of Aqueous Crude Leaf Extract of Senecio Biafrae Combined With Zinc on Testicular Function of Adult Male Sprague Dawley Rats. J Family Reprod Health. 2018 Mar;12(1):8-17. PMID: 30647753; PMCID: PMC6329991.
- Adetutu, W. A. Morgan, and O. Corcoran, "Ethnopharmacological survey and in vitro evaluation of wound-healing plants used in South-Western Nigeria," Journal of Ethnopharmacology, vol. 137, no. 1, pp. 50–56, 2011.
- Ajiboye B.O., Ibukun E.O., Edobor G., Ojo A.O. and Onikanni, S.A. 2013. Chemical composition of Senecio biafrae leaf. Scientific Journal of Biological Sciences, 2(8): 152-159.

- Ajiboye BO, Ojo OA, Okesola MA, Akinyemi AJ, Talabi JY, Idowu OT, Fadaka AO, Boligon AA, Anraku de Campos MM. In vitro antioxidant activities and inhibitory effects of phenolic extract of Senecio biafrae (Oliv and Hiern) against key enzymes linked with type II diabetes mellitus and Alzheimer's disease. Food Sci Nutr. 2018 Aug 13;6(7):1803-1810. doi: 10.1002/fsn3.749. PMID: 30349669; PMCID: PMC6189626.
- Aliero AA, Wara SH (2009) Validating the medicinal potential of Leptadenia hastata. Afr J Pharm Pharmacol 3: 335-338...
- American Association of Poison Control Centers. Practice Guideline: Acetaminophen poisoning: an evidence-based consensus guideline for out-of-hospital management. Clinical Toxicology. 2006: 44; 1-18.
- Antonia Mourtzikoua,, Maria Alepakia, Marilena Stamoulic,, Abraham Pouliakisa, Anastasios Sklirisc, Petros KarakitsosaEvaluation of serum levels of IL-6, TNF-α, IL-10, IL-2 and IL-4 in patients with chronic hepatitis. Inmunologia Vol. 33. Núm. 2. April June 2014 2014;33:41-50 DOI: 10.1016/j.inmuno.2014; 01.001
- Anusuya V., Sujatha G., Balaji G. L.(2020).
  Anti-oxidant, Antinflammation and Anti diabetic Activity of Novel Schiff Base of Derived from 2-Hydroxy benzoic acid (4-Chlorobenzylidine)-Hydrazide and 2-Hydroxybenzoic acid (3,4-Dimethoxybenzylidine)-Hydrazide. European Journal of Molecular & Clinical Medicine ISSN 2515-8260 Volume 07, Issue 9, 2020

- Aquino, R; Peluso, G; De Tommasi, N; De Simone, F; Pizza, C (1996). New polyoxypregnane ester derivative from Leptadenia hastata. J. Nat. Prdt. (59): 555-564
- Aruoma, Q.L. (2003). Methodological considerations for characterizing potential antioxidant actions of bioactive compounds. Plant foot mutat Res. 522(524), 9-20
- Atta, AH; Mouneir, SM (2005). Evaluation of somemedicinal plants extracts for antidiarrhoealactivity. Phytotheraph Research. (19): 481-485
- Awang-Kanak, F., Bakar, M. F. A., & Mohamed, M. Ethnobotanical note, total phenolic content, total flavonoid content, and antioxidative activities of wild edible vegetable, Crassocephalum crepidioides from Kota Belud, Sabah. IOP Conference Series: Earth and Environmental Science, 269(1): 12012 (2019).
- Ayoola MD, Adebajo AC, Zotor FB, Pinkoane MG. Justifying Antidiabetic Ethnomedicinal Claim of Senecio biafrae through Its Antihyperglycemic and Anti-Oxidant Activities. Ann Complement Altern Med. 2019; 1(2): 1006
- Bahar, E., Akter, KM., Lee, GH. et al. β-Cell protection and antidiabetic activities of Crassocephalum crepidioides (Asteraceae) Benth. S. Moore extract against alloxan-induced oxidative stress via regulation of apoptosis and reactive oxygen species (ROS). BMC Complement Altern Med 17, 179 (2017).
- Bello A, Aliero AA, Saidu Y, Muhammad S (2011) Hypoglycaemic and

- Hypolipidaemic Effects of Leptadeniahastata (Pers.) Decne in Alloxan Induced Diabetic Rats. Nigerian Journal of Basic and Applied Science 19: 187-192
- Bello, A., Aliero, A.A., Saidu, Y., Muhammad, S. (2011). Hypoglycaemic and Hypolipidaemic Effects of LeptadeniaHastata (pers.) Decne in Alloxan induces Diabetic Rats. The Nigerian Basic and Apllied science journal. 19(2), 187-192.
- Ben Menachem-Zidon O, Avital A, Ben-Menahem Y, Goshen I, Kreisel T, Shmueli EM, Segal M, Ben Hur T, Yirmiya R (Jul 2011). "Astrocytes support hippocampal-dependent memory and long-term potentiation via interleukin-1 signaling". Brain, Behavior, and Immunity. 25(5): 1008–16. doi:10.1016/j.bbi.2010.11.007. PMID 21093580.
- Betti, J.L., Yemefa'a, S.R.M., Nchembi, T.F. (2011). Contribution to the knowledge of no-woods forest products of the far North region of Cameroon. Journal of ecology and natural environment. 3(), 241-254.
- Boar, R.B. and Allen, J. (1973). B-Amyrintriterpenoids phytochemistry: 12, 2571-2578.
- Brocker C, Thompson D, Matsumoto A, Nebert DW, Vasiliou V (2010). "Evolutionary divergence and functions of the human interleukin (IL) gene family". Human Genomics. 5(1): 30–55. doi: 10.1186/1479-7364-5-1-30. PMC 3390169Freely accessible. PMID 21106488.

- Burkil, H.M. (2004). Brief description and detacts of over 4000 plants retrieved from www.aluka.org
- Chalasani NP, Hayashi PH, Bonkovsky HL, et al. ACG Clinical Guideline: The Diagnosis and Management of Idiosyncratic Drug-Induced Liver Injury. Am J Gastroenterol. 2014; 109: 950–966; published online 17 June 2014: PMID: 24935270 www.ncbi.nlm.nih.gov/pubmed/24935 270.
- Charls, C.D., Maribeth, C., Daniel, L.N., Kenneth, J.W. and David, A.B. (1996).Floral Gigantism in Refflesia Ceac.Science Express.
- Cherng, J.M., W. Chiang, J.H. Wang, C.M. Lin and C.M.Shin et al., 2008. of Anthraquinones edible wild vegetable Cassia tora stimulate proliferation of human CD4+ T lymphocytes and secretion of interferon-gamma or interleukin 10. Food Chem., 107: 1576-1580. DOI: 10.1016/j.foodchem.2007.10.005
- Chidume, F.C., H.O. Kwanashie, J.O. Adekeye, C. Wambebe and K.S. Gamaniel, 2002. Antinociceptive and smooth muscle contracting activities of the methanolic extract of Cassia tora leaf. J. Ethnopharmacol., 81: 205-209. DOI: 10.1016/S0378-8741(02)00079-X
- Cho, S.H., T.H. Kim, N.H. Lee, H.S. Son and I.J. Cho et al., 2005. Effects of Cassia tora fiber supplement on serum lipids in Korean diabetic patients. J. Med. Food, 8: 311-318. PMID: 16176140 Das, G., D. Ojha, B. Bhattacharya, M. Samanta and S. Ghosh et al., 2010. Evaluation of

- antimicrobial potentialities of leaves extract of the plant Cassia
- Danbatta, S.H., Aliyu, B.S. (2011). A survey of major Ethnomedical plant of Kano North Nigeria: their knowledge and uses by traditional healers. Bayero journal of pure and applied science. 4(): 28-34
- Das, K., Tiwari, R.K.S, Shrivastava, DK..

  Techniques for evaluation of medicinal plant products as antimicrobial Agent: current methods and future trends. Journal of Medicinal Plants Research. 2010: 4(2): 104-111.
- Dias, D.A., Urban, S., and Roessner, U. (2012).Ahistorical overview of natural product in Drug Discovery. Metabolites. 2(2). 303-336 Abbas A.B.; Lichtman A.H. (2009). "Ch.2 Innate Immunity". In Saunders (Elsevier). Basic Immunology. Functions and disorders of the immune system (3rd ed.). ISBN 978-1-4160-4688-2.
- Ferrero-Miliani L, Nielsen OH, Andersen PS, Girardin SE; Nielsen; Andersen; Girardin (February 2007). "Chronic inflammation: importance of NOD2 and NALP3 in interleukin-1beta generation". Clin. Exp. Immunol. 147 (2): 061127015327006—. doi:10.1111/j.1365-2249.2006.03261.x. PMC 1810472. PMID 17223962.
- Gaze DC (2007). "The role of existing and novel cardiac biomarkers for cardioprotection". Current Opinion in Investigational Drugs. 8(9): 711–7. PMID 17729182.

- Goldfrank LR, ed. Goldfrank's Toxicologic Emergencies. 8th ed. New York, NY: McGraw Hill; 2006.
- Granberg RA, Rasmuson AC (1999). "Solubility of paracetamol in pure solvents". Journal of Chemical & Engineering Data. 44 (6): 1391–95. doi: 10.1021/je990124v.
- Hayashi H, Mizuguchi H, Miyahara I, Nakajima Y, Hirotsu K, Kagamiyama H (2003). "Conformational change in aspartate aminotransferase on substrate binding induces strain in the catalytic group and enhances catalysis". J Biol Chem. 278 (11): 9481— 9488. doi:10.1074/jbc.M209235200. PMID 12488449.
- Hendrickson RG, McKeown, MJ.
  Acetaminophen. In: Marx JA,
  Hockberger RS, Walls RM, eds.
- Hoda Mohamed El-Emshaty, Wesam Ahmad Nasif,and Ibrahim Eldsoky Mohamed Serum Cytokine of IL-10 and IL-12 in Chronic Liver Disease: The Immune and Inflammatory Response. Dis Markers. 2015; 2015: 707254. Published online Kotowicz K, Callard RE, Friedrich K, Matthews DJ,
- Klein N (1996). "Biological activity of IL-4 and IL13 on human endothelial cells: functional evidence that both cytokines act through the same receptor". Int Immunol. 8 (12): 1915–25. doi:10.1093/intimm/8.12.1915. PMID 8982776.
- Olaniyan, Mathew. (2017). SCAVENGING ANTIOXIDATIVE BIOACTIVITIES

- OF Solanecio biafrae (Wòròwó) IN RABBITS INDUCED WITH ALCOHOL TOXICITY.
- Rosen's Emergency Medicine: Concepts and Clinical Practice. 8th ed. Philadelphia, PA: Elsevier Saunders; 2014:chap 148.
- Schippers, R.R. 2000. African indigenous vegetables. An overview of the cultivated species. Natural Resources Institute/ ACP-EU Technical Center for Agricultural and Rural Cooperation, Chatham, United Kingdom. pp. 214.
- Singh, P. P., Jha, S., & Irchhaiya, R. (2011).

  Antidiabetic and antioxidant activity of hydroxycinnamic acids from Calamintha Officinalis Moench.

  Medicinal Chemistry Research, 21(8), 1717–1721. doi:10.1007/s00044-011-9690-5
- Stevels, J.M.C. 1990. Traditional vegetables of Cameroon: an agrobotanical study. Wageningen Agricultural University Papers: No. 90-1. Wageningen University, Wageningen, Netherlands. pp. 262.
- Teoh NC, Chittun S, Farrell GC. Druginduced hepatitis. In: Feldman M, Friedman LS, Brandt LJ, eds. Sleisenger and Fordtran's Gastrointestinal and Liver Disease. 9th Philadelphia, PA: Elsevier Saunders; 2010: chap 86.